

Recent Progress In Bioavailability and Synergy of Curcumin, Quercetin, and Resveratrol



Michael Traub, ND, FABNO

plant intelligence.®
PROFESSIONAL RESOURCES

Introduction

Dietary supplementation of curcumin and other polyphenols confer many beneficial health effects, including prevention and treatment of cancer, cardiovascular disease, and diverse other inflammatory conditions. Efforts have been made to increase their poor bioavailability in order to increase their efficacy and lower the necessary dose.

Numerous preclinical and clinical trials have recently been undertaken to investigate the bioavailability and bioefficacy of polyphenolic compounds.¹ Major gains have been made to our knowledge of polyphenol absorption and metabolism. Various modifications to free polyphenolics (encapsulation, addition of piperine, nanoparticles, phospholipid carriers) and combinations of polyphenolics have been evaluated for their effects on absorption and efficacy. The challenges of assessing bioavailability and the concept of synergistic interactions between polyphenolics will be introduced. The purpose of this review is to place Lund and Mantuso's recent study of curcumin, quercetin and resveratrol² in the context of others to illuminate the current state of knowledge and guide further inquiry.

Bioavailability of polyphenolics

It is well recognized that polyphenolic compounds have relatively poor bioavailability. Lipinski's Rule of 5 states that compounds with five or more hydrogen bond donors (OH and NH groups), ten or more hydrogen bond acceptors (e.g. N and O), a molecular weight greater than 500, and Log P greater than 5 are usually poorly absorbed orally.^{3, 4}

In order for a polyphenol to achieve an effect *in vitro*, concentration can range >1000-fold, from <0.1 uM/L to >100 uM/L. However, physiologic concentrations do not exceed 10 uM/L, so the effects of polyphenols *in vitro* at concentrations in excess of this are generally not clinically valid, with the possible exception of the intestinal lumen.⁵

Some experiments of polyphenol bioavailability have been carried out using the Caco-2 cell culture model.⁶ Caco-2 human colon cancer cells have been widely used to predict

intestinal absorption of orally administered drugs and other compounds.⁷ Although derived from colon carcinoma, the cells differentiate when cultured under certain conditions to structurally and functionally resemble the enterocytes lining the small intestine. The *in vitro* apparent permeability (P-app) across Caco-2 monolayers correlates reliably with the *in vivo* fraction absorbed.

It is known that polyphenols interact non-covalently with proteins in plasma through hydrophilic or hydrophobic bonds, thus affecting bioavailability.⁸ Bioavailability of polyphenols is also significantly reduced by enzyme and microbial-mediated biotransformation and active efflux as well as by their physiochemical properties.⁹ The majority of polyphenols are not subject to Phase I hepatic detoxification due to their unfavorable structure for cytochrome P450 metabolism. Instead they are subject to direct Phase II metabolism (glucuronidation, sulfation and methylation). Females are more efficient in the glucuronidation of resveratrol.¹⁰ Polyphenols such as quercetin can modulate the intestinal microbiota and indirectly interfere with their own bioavailability.¹¹ Factors such as these are responsible for the unpredictable plasma levels of polyphenolic compounds seen following oral administration.

Curcumin

The biotransformation and pharmacokinetics of curcumin will serve as an example here. Curcumin is the yellow pigment in turmeric (*Curcuma longa*), curry powder, and mustard. It has poor solubility when ingested, and most is excreted in the feces and only small amounts of curcumin or its metabolites appear in blood. When absorbed, curcumin is quickly reduced in the intestine and conjugated in the liver to several glucuronide and sulfate products.¹² In human studies, doses of curcumin up to 180 mg have produced no detectable plasma levels.¹³ Doses up to 8 g have yielded only 0.5-2 uM.¹⁴ Curcumin concentration of 10 nmol/g tissue in human colorectal mucosa have been obtained with oral consumption of up to 3.6 g curcumin.¹⁵

Doses of curcumin up to 12 g/day have been shown to be safe in Phase I clinical trials.¹⁶ To achieve better bioavailability and reduce the dose of curcumin, various approaches have been taken. Piperine has been used as an adjuvant to inhibit glucuronidation. Liposomes, nanoparticles, phospholipid complexes, and structural analogues of curcumin (e.g., EF-24) have also been used to enhance absorption.¹⁷ An excellent review was recently published of nanotechnologies to enhance delivery of curcumin, quercetin and resveratrol for their anticancer effects *in vitro* and in mice.¹⁸

Quercetin

Quercetin is another example of a polyphenol with discrepancy observed between *in vitro* and *in vivo* studies attributed to absorption and metabolism. Quercetin (one of the most abundant natural flavonoids) is found at high concentrations in onions, apples, berries, red wine, broccoli, capers, pomegranate, and *Ginkgo biloba*. Although quercetin has been shown to possess effects including anti-inflammatory, antioxidant, anti-cancer, anti-anaphylaxis and anti-aging, it is unstable, has poor solubility and permeability, and low bioavailability. Approaches used to improve bioavailability of quercetin include liposomes, nanoparticles, inclusion complexes, and micelles.¹⁹

Resveratrol

Resveratrol, produced in berries, grapes, and peanuts in response to stress, injury, UV radiation and fungal infection,²⁰ is a third example of a polyphenolic compound with various beneficial effects. Resveratrol also is susceptible to oxidative decomposition, has poor solubility and low bioavailability, in large part due to extensive phase II metabolism.²¹ *In vitro* and animal studies have shown that resveratrol can act on a wide range of molecular targets, including sirtuins, influencing aging, transcription, apoptosis, inflammation, and stress resistance.²² Organic anion transporting polypeptides have recently been found in hamster ovary and breast cells to function as cellular uptake transporters for resveratrol and its major sulfates, and account for bioactivity.²³

The implication is clear that the proven biological effects of polyphenols *in vivo* do not correlate well with what we can observe about their bioavailability *in vitro*. This discrepancy has led to various strategies to improve their bioavailability and their bioactivity. Curcumin, quercetin and resveratrol will again serve as examples for how bioactivity can be improved.

Formulation approaches for improving bioavailability of polyphenols

Piperine

Piperine is the alkaloid responsible for the pungency of black pepper (*Piper nigrum*) and long pepper (*Piper longum*). Piperine strongly inhibits glucuronidation, affects the ultrastructure of intestinal brush border, and increases gastrointestinal transit time, thereby increasing absorption.^{24, 25, 26} In human volunteers, concomitant administration of 20 mg piperine with 2 g curcumin increased serum concentration by 2000% in the following 15 – 60 minutes ($p < 0.01$ at 0.25 h and $p < 0.001$ at 1 h).²⁷ The complete mechanism of piperine's bioavailability enhancement is not known. It does inhibit human CYP3A4 and the efflux pump P-glycoprotein biotransforming reactions in the liver.²⁸

A recent study demonstrated that piperine 20 mg alone and in combination with trans-resveratrol 200 mg significantly augmented cerebral blood flow (CBF) during task performance in comparison with placebo and resveratrol alone on separate days at least a week apart.²⁹ The plasma concentrations of resveratrol and its metabolites were not significantly different between the treatments, indicating that co-supplementation of piperine with resveratrol enhances the bioefficacy of resveratrol with regard to CBF effects, but not cognitive performance, and does this without altering bioavailability.

Lecithin, Phosphatidyl Choline, Proliposomes, Liposomes, and Phytosomes

Another method for enhancing bioavailability is complexing curcumin with a phospholipid, known as a phytosome. The phosphatidylcholine-curcumin complex is more readily incorporated into lipophilic cell membranes, making it significantly more bioavailable than unbound curcumin. One 500 mg dose of this complex is composed of 100 mg curcuminoids (ratio curcumin : demethoxycurcumin : bis-demethoxycurcumin 33 : 8 : 1), 200 mg soy lecithin and 200 mg microcrystalline cellulose. A randomized double-blind crossover trial of the complex and a standardized curcuminoid mixture showed curcuminoid absorption (plasma level) was about 29-fold higher for the complex.³⁰ However, only phase-2 metabolites were detected. Demethoxycurcumin, a more potent analogue in many *in vitro* anti-inflammatory assays, was the major plasma curcuminoid after administration of the complex.

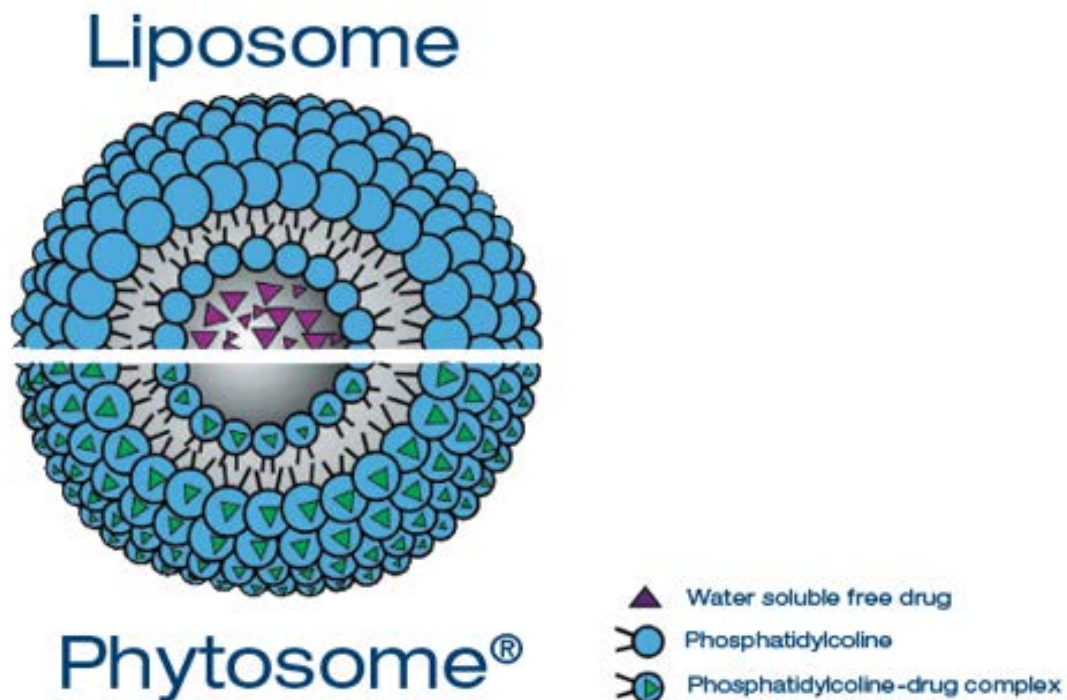


Fig. 1. Differences between a Phytosome[®] and a Liposome.

Although similar, phytosomes and liposomes have some basic differences (see Figure 1). The active ingredient in a liposome is dissolved in a central cavity, which limits its molecular interaction because of the surrounding phospholipid. In a phytosome, the active ingredient is dispersed in the phospholipid matrix (soy lecithin) and is integrated into the lipid membrane. Liposomes contain approximately five times the amount of phospholipids as phytosomes.³¹

Liposomes and phytosomes have been demonstrated to be excellent carriers for polyphenols, due to their biocompatibility, wide choice of physico-chemical properties and easy preparation.³² Conversion of polyphenols into phytosomes improves efficacy without compromising safety. Phytosome technology creates bonds between individual polyphenol molecules and one or more molecules of the phospholipid, phosphatidylcholine (PC). Molecular imaging reveals that PC molecule(s) enwrap each polyphenol; upon oral intake the amphipathic PC molecules probably escort the polyphenol through the intestinal epithelial cell membrane, and then into the bloodstream.³³

Aqueous solubility of quercetin improved by 12 fold (from 3.44 µg/ ml to 36.81 µg/ ml) in a quercetin-phospholipid complex, and bioactivity of quercetin was not adversely affected.³⁴

Proliposomes are powdered mixtures of drug and lipid, intended to produce multi-lamellar liposomes upon contact with aqueous media entrapping adequately hydrophobic drugs.³⁵ Proliposomes have distinct advantages over liposomes, as the free flowing powder can be incorporated into solid dosage forms such as capsules or tablets. The simple process of preparation makes them more suitable for industrial scale manufacturing: Multi-lamellar liposomes containing drug molecules are believed to be absorbed *via* lymphatic absorption. Based on lipid composition and type of lipid, there are two possible ways for absorption of entrapped drug. First, intact liposomes may be absorbed *via* endocytosis by enterocytes. Second, due to their considerable size, the absorbed liposomes may be transported *via* the lymphatic system into systemic

circulation. Hence proliposomes can be a solution to issues related to solubility, permeability, P-glycoprotein efflux, and hepatic metabolism of polyphenols.³⁶

A Chinese group developed a novel curcumin-phytosome-loaded chitosan microspheres (Cur-PS-CMs) delivery system and showed that the *in vitro* release rate of curcumin from it was slower than that from curcumin-loaded chitosan microspheres.³⁷ Rats dosed with Cur-PS-CMs had a 1.67- and a 1.07-fold enhancement of curcumin absorption compared with Cur-PS and Cur-CMs respectively. Half-life from Cur-PS-CMs (3.16h) was longer than that of Cur-PS (1.73h) and Cur-CMs (2.34h).

A proliposomal formulation of resveratrol resulted in significantly improved rate and extent of absorption of unmetabolized resveratrol *in vitro* and *in vivo*. Area under the curve and maximum concentration were twofold higher than plain resveratrol.³⁸

Nanotechnology

Nanoparticles (NPs) range from 1 to 100 nm in size and are composed of biodegradable and biocompatible polymers including starch, co-poly(lactic acid/glycolic acid) (PLGA), and poly(lactic acid) (PLA). NPs can be useful carriers for polyphenols in that they permit encapsulation of single or multiple compounds in their core or on their surface, and the time and rate of polymer degradation and subsequent delivery of the polyphenol can be controlled. Khushnud and Mousa recently reviewed studies published within the past 5 years on the use of NPs to enhance polyphenol delivery.³⁹

Human serum albumin (HAS) NPs are non-toxic and immunogenic, and allow endothelial transcytosis of unbound and albumin-bound plasma constituents to the extravascular space. Mice with multiple melanomas were injected with both free and HSA encapsulated curcumin.⁴⁰ Nanoparticle-encapsulated curcumin concentrations were 14 times greater in melanomas compared to free curcumin. Xenograft mouse models were also used to test antitumor activity on human colon cancer (HCT116) and human pancreatic carcinoma (MiaPaCa2). Nanoparticle-bound curcumin inhibited tumor growth more (50%) for both cell lines compared to free curcumin (18%).

A Japanese group developed a curcumin formulation dispersed with colloidal nanoparticles (Oblate) in 2011.⁴¹ They found the AUC after oral administration to rats was greater than 40-fold higher than normal curcumin powder. Healthy human volunteers (8 males and 6 females) were then randomly assigned to 30 mg of the formulation or curcumin powder in 100 mL water. The AUC of the formulation was 27-fold higher than curcumin powder. They also found the 30 mg dose of the formulation in mineral water after alcohol ingestion significantly reduced acetaldehyde blood concentration versus mineral water only in a cross-over trial of 7 males. Acetaldehyde is considered to be responsible for the headache and other hangover symptoms associated with excessive alcohol drinking.

Conventional methods for the synthesis of biodegradable NPs rely on toxic molecules such as polyvinyl alcohol, polyethylene glycol, D-alpha-tocopheryl poly(ethylene glycol 1000) succinate as stabilizers/emulsifiers. A green approach uses plant extract synthesized PLA NPs for the sustained release of quercetin.⁴²

A safe, self-nanoemulsifying lipid based delivery system of resveratrol has been shown to significantly increase *in vitro* cytotoxicity in MCF-7 breast cancer cells and increased anti-angiogenic activity *in vivo* in chick chorioallantoic membrane assays.⁴³ Resveratrol-loaded carboxymethyl chitosan NPs improve antioxidant activity *in vitro*, and in rats increase absorption and relative bioavailability by 3.516 times more than raw resveratrol.⁴⁴

Synergistic interactions of polyphenolics: more than the sum of its parts

The concept of synergy, that a whole or purified extract of a plant confers advantages over an isolated constituent, including the efficacy of low doses of herbs in an efficacious botanical formula, is a fundamental philosophical underpinning of the major traditions of phytomedicine including Western herbalism, traditional Chinese medicine and Ayurveda. Evidence supporting synergism in herbal medicine has been reviewed by Williamson⁴⁵

and Wagner and Ulrich-Merzenich.^{46,47} Growing sophistication in analytical chemistry, and molecular, genomic, and biological methods has helped elucidate a variety of synergistic mechanisms of phytochemicals. Synergy is observed when constituents of an extract have multiple target mechanisms of action, and as we have seen above, are formulated to enhance bioavailability. The evolving rationale of synergy has led to greater legitimacy for phytotherapy.

Synergy is not always straightforward. A number of theoretical possibilities exist, that may or may not be seen in clinical practice, and may be unpredictable. It is possible that synergy can occur at one dose of a combination of substances and antagonism at another. Other potential complicating factors are the presence of unstable constituents, length of administration, microflora metabolism, host factors such as pH, and unknown active constituents.

A team of Dutch authors recently reviewed the mechanisms of combined action of chemopreventive dietary compounds.⁴⁸ These agents can show significant activity at concentrations where any single one agent is inactive, suggesting synergy. Although mechanistic understanding is limited, numerous combinations of complementary modes of action may be at play. Evidence is mounting that the activity of phytochemicals administered as dietary supplements alone do not explain observed health benefits of a diet rich in fruits, vegetables, spices, tea, coffee and wine.

Comparative bioavailability and synergy of polyphenol formulations

Differences in study design, subjects, analytical methods, and administration significantly limit comparisons between different polyphenol formulations. Here we shall examine three studies that have made comparative measurements of different formulations of polyphenols.

The first study was a pilot cross-over design of 11 healthy volunteers that compared 2000 mg of a patented formulation, with equivalent doses of a combination of curcumin-

lecithin-piperine, and normal curcumin.⁴⁹ The formula content in blood measured by HPLC at 1, 2, 3, 4, 5, 6 and 8 h post-drug was 6.93-fold greater in the AUC than normal curcumin and 6.37-fold compared to the curcumin-lecithin-piperine formula.

The second study focused on comparative increases in relative absorption of curcuminoids (curcumin, demethoxycurcumin, bisdemethoxycurcumin) and the metabolite tetra- hydrocurcumin after oral administration of three different curcumin formulations in comparison to a standardized curcumin mixture of 1,800 total curcuminoids (CS).⁵⁰ The three formulations were equivalent doses (376 mg total curcuminoids) of a curcumin phytosome (CP), curcumin with volatile oils of turmeric rhizome (CTR), and a novel formulation of curcumin with a water soluble carrier (polyvinyl pyrrolidone) and tocopherol and ascorbyl palmitate as antioxidants) (CHC). 12 healthy volunteers completed this randomized, double-blind, crossover study, consisting of 4 trials with 9 blood draws each, separated by at least 7 days. Samples were analyzed by HPLC-MS/MS. Total curcuminoids appearance in the blood was 1.3-fold higher for CTR and 7.9-fold higher for CP in comparison to unformulated CS. CHC showed a 45.9-fold higher absorption over CS and significantly improved absorption over CP (5.8-fold) and CTR (34.9-fold, all $p < 0.001$).

The third study, a recent *in vitro* epithelial model of absorption of various combinations of quercetin (Q), resveratrol (R), and curcumin (C), provides preliminary evidence of the respective bioavailability of these three bioactive polyphenols across a caco-2 monolayer and how they can act synergistically.² The three polyphenols were quantified using an Alliance HPLC system (Waters Corporation, MA). The apparent permeability coefficient (P_{app} , cm/s) was calculated for each compound/combination using the following equation:

$P_{app} = (dC_b/dT)/(C_a \cdot A)$ where (dC_b/dT) is the rate of appearance of the constituent on the basal side (nmol/s), C_a is the concentration of the constituent on the apical side (50 μ M) and A is the area of the monolayer (0.3 cm^2). The P_{app} , cm/s describes the ability of individual compounds to pass through the cell monolayer under the experimental

conditions.

The molecular structure of quercetin (Q), resveratrol (R) and curcumin (C) share similar characteristics that suggest they may have similar routes of uptake and efflux and thus may affect each other's absorption. The authors hypothesized that combinations of the three compounds, with and without piperine (P), 200 uM, would therefore affect their apical-to-basal absorption. Q, R and C were applied apically in isolation or in combination at 50 uM and measured in the basal chamber at 30 minutes.

Resveratrol had the largest enhancement in permeability when combined with Q (310%), and then with C (300%). The combination of Q & C increased the permeability of R by 323% (from 7.70 \pm 1.10 to 25.09 \pm 0.48). The combination of Q, C, and P increased permeability the most (350%; 27.22 \pm 0.89).

In isolation, Q had the highest permeability (10.99 \pm 0.77), R had 7.77 \pm 1.10, and C had the poorest (0.99 \pm 0.11).

Curcumin had 147% increased permeability when combined with Q, and 188% increased permeability when combine with Q & R. Addition of P resulted in the greatest increase: 229%.

Quercetin permeability was not significantly enhanced with any combination, but was maximal when combined with R: (11.90 \pm 0.52) and worst when combined with R, C and P (8.89 \pm 0.40).

Piperine appeared to enhance C and R and adversely affect Q permeability.

These results suggest that delivering these compounds in combination may improve the acute absorption of curcumin and resveratrol compared to supplementation with single polyphenol, reducing the need for higher doses and simplifying treatment with combination formulas.

The authors acknowledge that earlier investigations with longer exposures to polyphenols have shown changes in expression of UGT1A1, CYP3A4 and P-gp.^{51 52} Therefore, long term use of these combinations may further affect their absorption profiles.

Finally, it is important to consider that the caco-2 model is a valid model of absorption but not necessarily bioavailability. Also, the caco-2 cells used in different laboratories around the world have diverged significantly, which makes it difficult to compare results across labs.⁵³

This study strongly calls for an *in vivo* study to assess whether the *in vitro* findings are similar. A crossover human trial measuring absorption of the 3 polyphenols separately and in combination should be performed.

Bioavailability of parent compounds vs. select metabolites

The data on the bioavailability of polyphenols presented above considered the presence of intact polyphenols *in vitro* or in the blood, i.e. the ingested compounds or their conjugates. The trillions of intestinal microflora also play a significant role in the metabolism of polyphenols. After microbial enzyme-catalyzed deconjugation of any polyphenol conjugates that reach the colon, there are 2 possible routes available; (1) absorption of the intact polyphenol through the intestinal lining and passage into the bloodstream (as free or conjugated forms) or biotransformation of the original polyphenol into metabolites. The absorption data presented above include the contribution of the absorption of intact polyphenols in the colon but do not include the breakdown contribution. Microbial metabolism of polyphenols can break them down into simpler phenolic compounds that are common to many different polyphenols, some of which could have unique biological effects. For example, *Lactobacillus plantarum* IFPL935 has been shown to significantly increase the concentration of the catabolites of red wine polyphenols.⁵⁴ Some of these microbial-derived metabolites may be responsible for the effects attributed to resveratrol.

Health benefits associated with curcumin may depend on the amount of certain curcuminoids in the various available formulations. Further studies are needed to determine which curcuminoids are most potent and what doses are most effective for clinical applications.

Conclusion

Curcumin is a well-studied down-regulator of the inhibitory kappa B alpha kinase, a key activator of nuclear-factor kappa B.⁵⁵ Blocking this central mediator makes curcumin therapeutically active in various inflammatory conditions including arthritis, cardiovascular disease, metabolic disease, neurodegeneration and tumorigenesis.

Curcumin may work synergistically and have improved bioavailability when combined with other polyphenolics and phytochemicals. For example, for cardiovascular applications, quercetin, resveratrol, piperine, hawthorn berry, coleus forskohlii root, and ginger root may be considered. For musculoskeletal pain and health, quercetin, devil's claw root, boswellia gum resin, piperine, ginger root, rosemary leaf, wild lettuce, Jamaican dogwood bark, feverfew leaf and flower would be options. For respiratory support, quercetin, stinging nettle leaf, feverfew leaf and flower, goldenseal root, piperine and ginger root are considerations.

Curcumin's diverse array of molecular targets affords it great potential as a therapeutic agent. There is intense interest in its therapeutic potential as evidenced by the number of ongoing phase II and III clinical trials. The primary obstacle to utilizing curcumin therapeutically has been its limited systemic bioavailability, but there are studies suggesting the various curcumin formulations may make it more effective and better absorbed. Results from completed clinical trials are encouraging and trials currently being conducted for both inflammatory conditions and cancer should clarify curcumin's value as a therapeutic agent and confirm some of the mechanisms responsible for its efficacy.

The human pharmacokinetic data on bioavailability of polyphenols are incomplete, which

limits clinical research and product development. The synergistic effects of polyphenolics should be further explored for additional beneficial and reliable outcomes in the prevention and treatment of inflammatory conditions, including cancer and cardiovascular disease.

Author

Michael Traub, ND, DHANP, CCH, FABNO attended the University of California, Irvine where he received a B.S. in biological sciences, and graduated from National College of Naturopathic Medicine (NCNM) in 1981. He completed a residency program in Family Practice and Homeopathy at NCNM. Since 1986, Dr. Traub has been Medical Director of an integrative healthcare center (Lokahi Health Center) in Kailua Kona, Hawaii. He is board certified in naturopathic oncology as well as homeopathy. Dr. Traub has conducted a wide range of research studies on homeopathic growth factors in HIV and AIDS, Attention Deficit and Hyperactivity Disorder, Paroxysmal Nocturnal Hemoglobinuria in Bone Marrow Failure Syndromes, Hypovitaminosis D, and Elderberry Extract for Prevention of Influenza.

He is the author of “Essentials of Dermatological Diagnosis and Natural Therapeutics” and “Essentials of Dermatological Diagnosis and Integrative Therapeutics.” He has contributed to several textbooks including the Textbook of Natural Medicine. He serves on the editorial board of the Natural Medicine Journal, the International Journal of Naturopathic Medicine, and Holistic Primary Care. He is a member of the Board of Directors of the Integrated Healthcare Policy Consortium and was co-author of the “Final Report of the National Policy Dialogue to Advance Integrated Health Care: Finding Common Ground, 2001-2002, and co-editor of “The Affordable Care Act & Beyond: A Stakeholder Conference on Integrated Healthcare Reform,” Sept. 2010.

He currently serves on the Scientific Advisory Boards for Gaia Herbs, Inc., Kamedis, and Nordic Naturals.

References

- ¹<http://clinicaltrials.gov/ct2/show/NCT01288859?term=bioavailability+AND+curcumin&rank=4>
- ² Lund KC, Pantuso T. Combination Effects of Quercetin, Resveratrol and Curcumin on *in vitro* intestinal absorption. *J Rest Med* 2014;3:112-120.
- ³ Yang CS, Sang S, Lambert JD, Lee M. Bioavailability issues in studying the health effects of plant polyphenolic compounds. *Mol Nutr Food Res*. 2008, 52:S139-S151.
- ⁴ Lipinski JD, Lombardo F, Dominy BW, Feeney PJ. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv Drug Deliv Rev*. 2001, 46:3-26.
- ⁵ Williamson G, Manach C. Bioavailability and bioefficacy of polyphenols in humans. II Review of 93 intervention studies. *Am J Clin Nutr*. 2005;81(suppl):243S–55S
- ⁶ Zhang L., Chow MS, Zuo, Z. Effect of the co-occurring components from green tea on the intestinal absorption and disposition of green tea polyphenols in Caco-2 monolayer model. *J. Pharm. Pharmacol*. 2006, 58:37-44.
- ⁷ Artursson P, Palm K, Luthman K. Caco-2 monolayers in experimental and theoretical predictions of drug transport. *Adv. Drug Deliv. Rev*. 2001, 46, 27-43.
- ⁸ Xiao J. Polyphenol-plasma proteins interaction; its nature, analytical techniques, and influence on bioactivities of polyphenols. *Curr Drug Metab*. 2013 May;14 (4):367-8
- ⁹ Manach C, Williamson G, Morand C, Scalbert A. et al., Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *Am. J. Clin. Nutr*. 2005, 81:230S-242S
- ¹⁰ Dellinger RW, Garcia AM, Meyskens FL Jr. Differences in the glucuronidation of resveratrol and pterostilbene: altered enzyme specificity and potential gender differences. *Drug Metab Pharmacokinet*. 2014;29(2):112-9.
- ¹¹ Duda-Chodak A. The inhibitory effect of polyphenols on human gut microbiota. *J Physiol Pharmacol*. 2012 Oct;63(5):497-503.
- ¹² Ireson CR, Jones DJ, Orr S, Coughtrie MW, et al. Metabolism of the cancer chemopreventive agent curcumin in human and rat intestine. *Cancer Epidemiol. Biomarkers Prev*. 2002,11:105-111.
- ¹³ Sharma RA, McLelland HR, Hill KA, Ireson, CR, et al., Pharmacodynamic and pharmacokinetic study of oral Curcuma extract in patients with colorectal cancer. *Clin. Cancer Res*. 2001, 7:1894-1900
- ¹⁴ Cheng AL, Hsu CH, Lin JK, Hsu MM, et al. Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions. *Anticancer Res*. 2001, 21:2895-2900.
- ¹⁵ Garcea G, Jones DJ, Singh R, Dennison AR, et al. Detection of curcumin and its metabolites in hepatic tissue and portal blood of patients following oral administration. *Br. J. Cancer* 2004, 90:1011-1015.
- ¹⁶ Lao CD, Ruffin MT, Normolle D, Heath DD, Murray SI, Bailey JM, Boggs ME, Crowell J, Rock CL, Brenner DE: Dose escalation of a curcuminoid formulation. *BMC Complement Altern Med* 2006, 6:10.
- ¹⁷ Anand P, Kunnumakkara AB, Newman RA, Aggarwal BB. Bioavailability of curcumin: problems and promises. *Mol Pharm*. 2007 Nov-Dec;4(6):807-18.
- ¹⁸ Aras A, Khokhar AR, Qureshi MZ, et al. Targeting Cancer with Nano-Bullets: Curcumin, EGCG, Resveratrol and Quercetin on Flying Carpets. *Asian Pac J Cancer Prev*. 2014; 15(9):3865-3871
- ¹⁹ Cai X, Fang Z, Dou J, Yu A, Zhai G. Bioavailability of quercetin: problems and promises. *Curr Med Chem*. 2013;20(20):2572-82.
- ²⁰ Hammerschmidt R. PHYTOALEXINS: What Have We Learned After 60 Years? *Annu Rev Phytopathol* 1999;37:285–306.
- ²¹ Francioso A, Mastromarino P, Masci A, d'Erme M, Mosca L. Chemistry, stability and bioavailability of resveratrol. *Med Chem*. 2014 May;10(3):237-45.
- ²² Preyat N, Leo O. Sirtuin deacylases: a molecular link between metabolism and immunity. *J. Leuk. Biol*. 2013. 93 (5):669–680.
- ²³ Riha J, Brenner S, Böhmendorfer M, Giessrigl B, Pignitter M, Schueller K, Thalhammer T, Stieger B, Somoza V, Szekeres T, Jäger W. Resveratrol and its major sulfated conjugates are substrates of organic anion transporting polypeptides (OATPs): Impact on growth of ZR-75-1 breast cancer cells. *Mol Nutr Food Res*. 2014 Jul 3. doi: 10.1002/mnfr.201400095. [Epub ahead of print]
- ²⁴ Bajad S, Bedi KL, Singla AK, Johri RK. Piperine inhibits gastric emptying and gastrointestinal transit in

rats and mice. *Planta Med.* 2001, 67: 176-179.

²⁵ Reen RK, Jamwal DS, Taneja SC, Koul JL, et al. Impairment of UDP-glucose dehydrogenase and glucuronidation activities in liver and small intestine of rat and guinea pig in vitro by piperine. *Biochem. Pharmacol.* 1993, 46: 229-238

²⁶ Srinivasan K. Black pepper and its pungent principle – piperine: a review of diverse physiological effects. *Crit Rev Food Sci Nutr.* 2007. 47(8):735-748.

²⁷ Shoba G, Joy D, Joseph T, Majeed M, Rajendran R, Srinivas PS. Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Med.* 1998 May;64(4):353-6.

²⁸ Bhardwaj RK, Glaeser H, Becquemont L, et al. Piperine, a major constituent of black pepper, inhibits human P-glycoprotein and CYP3A4. *J Pharmacol Exp Ther.* 2002.302(2):645-50.

²⁹ Wightman EL, Reay JL, Haskell CF, Williamson G, Dew TP, Kennedy DO. Effects of resveratrol alone or in combination with piperine on cerebral blood flow parameters and cognitive performance in human subjects: a randomised, double-blind, placebo-controlled, cross-over investigation. *Br J Nutr.* 2014 Jul;112(2):203-13.

³⁰ Cuomo J, Appendino G, Dern AS, Schneider E, McKinnon TP, Brown MJ, Togni S, Dixon BM. Comparative absorption of a standardized curcuminoid mixture and its lecithin formulation. *J Nat Prod.* 2011 Apr 25;74(4):664-9.

³¹ Semalty A, Semalty M, Rawat MS, Franceschi F. Supramolecular phospholipids-polyphenolics interactions: the PHYTOSOME strategy to improve the bioavailability of phytochemicals. *Fitoterapia.* 2010 Jul;81(5):306-14

³² Bonechi C, Martini S, Ciani L, Lamponi S, Rebmann H, Rossi C, Ristori S. Using liposomes as carriers for polyphenolic compounds: the case of trans-resveratrol, *PLoS One.* 2012;7(8):e41438.

³³ Kidd PM. Bioavailability and activity of phytosome complexes from botanical polyphenols: the silymarin, curcumin, green tea, and grape seed extracts. *Altern Med Rev.* 2009 Sep;14(3):226-46.

³⁴ Singh D, Rawat MS, Semalty A, Semalty M. Quercetin-phospholipid complex: an amorphous pharmaceutical system in herbal drug delivery. *Curr Drug Discov Technol.* 2012 Mar;9(1):17-24.

³⁵ Shaji J, Bhatia V. Proliposomes: a brief overview of novel delivery system. 2013. *Int J Pharm Biosci.* 4:150–160

³⁶ Basavaraj S, Betageri GV. Can formulation and drug delivery reduce attrition during drug discovery and development – review of feasibility, benefits and challenges. *Acta Pharm Sinica B* 2014. 4(1):3-17.

³⁷ Zhang J, Tang Q, Xu X, Li N. Development and evaluation of a novel phytosome-loaded chitosan microsphere system for curcumin delivery. *Int J Pharm* 2013.44891):168-74.

³⁸ Basavaraj S, Betageri GV. Improved oral delivery of resveratrol using proliposomal formulation: investigation of various factors contributing to prolonged absorption of unmetabolized resveratrol. *Expert Opin Drug Deliv.* 2014 Apr;11(4):493-503.

³⁹ Khushnud T, Mousa SA. Potential role of naturally derived polyphenols and their nanotechnology delivery in cancer. *Mol Biotechnol.* 2013 Sep;55(1):78-86.

⁴⁰ Kim TH, Jiang HH, Youn YS, Park CW, Tak KK, Lee S, et al. Preparation and characterization of water-soluble albumin-bound curcumin nanoparticles with improved antitumor activity. *Int J Pharm.* 2011.403:285–291.

⁴¹ Sasaki H, Sunagawa Y, Takahashi K, Imaizumi A, Fukuda H, Hashimoto T, Wada H, Katanasaka Y, Kakeya H, Fujita M, Hasegawa K, Morimoto T. Innovative preparation of curcumin for improved oral bioavailability. *Biol Pharm Bull* 2011, 34(5):660–665.

⁴² Kumari A, Kumar V, Yadav SK. Plant extract synthesized PLA nanoparticles for controlled and sustained release of quercetin: a green approach. *PLoS One.* 2012;7(7):e41230.

⁴³ Pund S, Thakur R, More U, Joshi A. Lipid based nanoemulsifying resveratrol for improved physicochemical characteristics, in vitro cytotoxicity and in vivo antiangiogenic efficacy. *Colloids Surf B Biointerfaces.* 2014 Aug 1;120:110-7

⁴⁴ Zu Y, Zhang Y, Wang W, Zhao X, Han X, Wang K, Ge Y. Preparation and in vitro/in vivo evaluation of resveratrol-loaded carboxymethyl chitosan nanoparticles. *Drug Deliv.* 2014 Jun 11:1-11.

⁴⁵ Williamson EM. Synergy and other interactions in phytomedicines. *Phytomedicine* 2001. 8(5):401-409.

⁴⁶ Wagner H, Ulrich-Merzenich G. Synergy research: approaching a new generation of phytopharmaceuticals. *Phytomedicine* 2009.16:97-110.

⁴⁷ Wagner H. Synergy research: approaching a new generation of phytopharmaceuticals. *Fitoterapia* 2011. 82(1):34-7.

-
- ⁴⁸ de Kok TM, van Breda SG, Manson MM. Mechanisms of combined action of different chemopreventive dietary compounds. *Eur J Nutr* 2008;47 (Suppl 2):51-59.
- ⁴⁹ Antony B, Merina B, Iyer VS, Judy N, Lennertz K, Joyal S. *Indian J Pharm Sci* 2008 70(4):445-449
- ⁵⁰ Jäger R, Lowery RP, Calvanese AV, Joy JM, Purpura M, Wilson JM. Comparative absorption of curcumin formulations. *Nutr J*. 2014; 13: 11.
- ⁵¹ Iwuchukwu OF, Tallarida RJ, Nagar S. Resveratrol in combination with other dietary polyphenols concomitantly enhances antiproliferation and UGT1A1 induction in Caco-2 cells. *Life Sci*. 2011;88(23-24):1047-54.
- ⁵² Okura T, Ibe M, Umegaki K, Shinozuka K, Yamada S. Effects of dietary ingredients on function and expression of P-glycoprotein in human intestinal epithelial cells. *Biol Pharm Bull*. 2010;33(2):255-9.
- ⁵³ Sambuy Y, Angelis I, Ranaldi G, Scarino M, Stamatii A, Zucco F. The caco-2 cell line as a model of the intestinal barrier: influence of cell and culture-related factors on caco-2 cell functional characteristics. *Cell Biol Toxicol* 2005. 21(1):1-26
- ⁵⁴ Barroso E, Sánchez-Patán F, Martín-Alvarez PJ, Bartolomé B, Moreno-Arribas MV, Peláez C, Requena T, van de Wiele T, Martínez-Cuesta MC. *Lactobacillus plantarum* IFPL935 favors the initial metabolism of red wine polyphenols when added to a colonic microbiota. *J Agric Food Chem*. 2013 Oct 23;61(42):10163-72.
- ⁵⁵ Jobin C, Bradham CA, Russo MP, Juma B, Narula AS, Brenner DA, Sartor RB. Curcumin blocks cytokine-mediated NF-kappa B activation and proinflammatory gene expression by inhibiting inhibitory factor I-kappa B kinase activity. *J Immunol*. 1999 Sep 15;163(6):3474-83.